



Requisition for Molecular Diagnostic Tests

PATIENT LAST: _____ FIRST: _____ Date of Request: _____
 Gender: _____ Birth Date: _____ CRIS Order # (if any): _____
 Pathology Case Number: _____ Medical Record Number: _____
 LP Resident/Fellow: Dr. _____ Requesting Physician: Dr. _____ Telephone #: _____

Specimen Information

Formalin-fixed, Paraffin-embedded Tissue (see #5 on back page for unstained slide ordering code):

Slides:	NIH recut	Outside Unstained Slides		
Involvement:	Full	Partial (circle area of interest)	Tumor Content:	%
Outside Case #:	_____	SoftPath Block ID: _____	Tissue Source: _____	_____
	_____	_____	_____	_____
	_____	_____	_____	_____
Other (specify):	_____	Tissue Source: _____	_____	_____

Clinical Information

Diagnosis / Preliminary Diagnosis: _____
 Brief Clinical History: _____
 Molecular History (MD#, DNA#, molecular result): _____

Tests Requested (If microarrays is high priority for classification, please check the check box. Otherwise, NGS will be run first)

Hematopathology	Sarcoma	COMPASS	High Priority
B Cell Clonality: IGH@ IGK@	Desmoplastic Round Cell: EWSR1/WT1, t(11;22)	DNA Methylation Microarrays Classifier TruSight Oncology 500 (TSO500)	
T Cell Clonality: TRG@	Ewings: EWSR1/ERG, t(21;22) EWSR1/FLI1 Type1, t(11;22) EWSR1/FLI1 Type2, t(11;22)		
Tumor-associated Virus:	Rhabdomyosarcoma: PAX3/FOXO1, t(2;13) PAX7/FOXO1, t(1;13)	Single Mutation Test	
EBV (EBNA2) HHV8 HTLV-I (Pol)	Synovial: SYT/SSX1, t(X;18) SYT/SSX2, t(X;18)	BRAF V600E EGFR T790M MYD88 L265P TERT Promoter (C228T & C250T)	
		DNA Extraction for DLM Microbiology Lab	

MD #: _____	Date Accession: _____
DNA #: _____	Concentration: _____ (ng/ul) Vol. _____ (ul) Quality: _____
RNA #: _____	Concentration: _____ (ng/ul) Vol. _____ (ul) Quality: _____
<u>Test Name</u>	<u>Test Result</u>
_____	_____
_____	_____
_____	_____
_____	_____
_____	_____
_____	_____
Signature: _____ Date: _____	

Instructions for Specimen Submission of Molecular Diagnostic Tests

1. Please type or clearly print all information and check appropriate boxes applied to your specimens.
2. Paraffin-embedded tissue should be formalin fixed. DNA extracted from tissues fixed in B-5 or other heavy metal containing fixatives does not amplify well.
3. For B and T-cell clonality assays and single mutation tests, the minimal amount of tissue required is 1 cm².

$$1 \text{ cm}^2 = \square \text{ or } \bigcirc$$

4. For all NGS assays, please submit 1 H&E slide and 10 unstained slides for small biopsies and cytology specimens, or 5 unstained slides for larger excisional biopsies and resection specimens.
5. Unstained slide ordering codes in SoftPath:
Hematopathology, BRAF V600E, MYD88 - **UNST** Sarcoma,
TERT Promoter, Methylation Classifier, TSO500 - **UNMOL**
Pulling archived unstained slides - **YMOL**
6. Please include the Outside Case #, SoftPath Block ID, and Tissue Source on your request. This information is required for QM/QA in the lab and during the molecular sign-out.
7. Always submit the completed requisition form with the specimen. For cases from the Clinical Center, you must place a CRIS order as well.
8. **Please provide a molecular diagnostic history with LP MD# if a prior molecular test was performed in house. To compare B & T cell clonality between the current case and a previous case that was signed out before 6/1/2009 (gel detection method), please submit unstained slides from the previous case for reanalysis with our current CE detection method.**

Contacts:

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